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## Fluxes of nitrous oxide and methane in different coastal Suaeda salsa marshes of the Yellow River estuary, China

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## HIGHLIGHTS

- ▶ Fluxes of N<sub>2</sub>O and CH<sub>4</sub> in coastal marsh had different spatial and temporal variations.
- $\blacktriangleright$  Coastal marsh represented N<sub>2</sub>O emission and CH<sub>4</sub> sink during sampling campaigns.
- ▶ N<sub>2</sub>O comprised the principal part of total CO<sub>2</sub>-e emissions during spring and winter.
- ► Contributions of CH<sub>4</sub> to total CO<sub>2</sub>-e could not be ignored during summer and autumn.

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## ABSTRACT

The spatial and temporal variations of the fluxes of nitrous oxide (N<sub>2</sub>O) and methane (CH<sub>4</sub>) and associated abiotic sediment parameters were quantified for the first time across the coastal marsh dominated by Suaeda salsa in the Yellow River estuary during 2009/2010. During all times of day and the seasons measured, N<sub>2</sub>O and CH<sub>4</sub> fluxes from coastal marsh ranged from  $-0.0147 \text{ mgN}_2\text{O} \text{ m}^{-2} \text{ h}^{-1}$  to 0.0982 mgN<sub>2</sub>.  $0 \text{ m}^{-2} \text{ h}^{-1}$  and  $-0.7421 \text{ mgCH}_4 \text{ m}^{-2} \text{ h}^{-1}$  to  $0.4242 \text{ mgCH}_4 \text{ m}^{-2} \text{ h}^{-1}$ , respectively. The mean N<sub>2</sub>O fluxes in spring, summer, autumn and winter were  $0.0325 \text{ mgN}_20 \text{ m}^{-2} \text{ h}^{-1}$ ,  $0.0089 \text{ mgN}_20 \text{ m}^{-2} \text{ h}^{-1}$ ,  $0.0119 \text{ mgN}_20 \text{ m}^{-2} \text{ h}^{-1}$  and  $0.0140 \text{ mgN}_20 \text{ m}^{-2} \text{ h}^{-1}$ , and the average CH<sub>4</sub> fluxes were  $-0.0109 \text{ mgCH}_4$   $m^{-2} h^{-1}$ , -0.0174 mgCH<sub>4</sub>  $m^{-2} h^{-1}$ , -0.0141 mgCH<sub>4</sub>  $m^{-2} h^{-1}$  and -0.0089 mgCH<sub>4</sub>  $m^{-2} h^{-1}$ , respectively, indicating that the coastal marsh acted as N<sub>2</sub>O source and CH<sub>4</sub> sink. Both N<sub>2</sub>O and CH<sub>4</sub> fluxes differed significantly between times of day of sampling. N<sub>2</sub>O fluxes differed significantly between sampling seasons as well as between sampling positions, while CH<sub>4</sub> fluxes had no significant differences between seasons or positions. Temporal variations of  $N_2O$  emissions were probably related to the effects of vegetation (S. salsa) during summer and autumn and the frequent freeze/thaw cycle of sediment during spring and winter, while those of CH<sub>4</sub> fluxes were controlled by the interactions of thermal conditions and other abiotic factors (soil moisture and salinity). Spatial variations of  $N_2O$  and  $CH_4$  fluxes were primarily affected by soil moisture fluctuation derived from astronomic tide, sediment substrate and vegetation composition. N<sub>2</sub>O and CH<sub>4</sub> fluxes, expressed as CO<sub>2</sub>-equivaltent (CO<sub>2</sub>-e) emissions, showed that N<sub>2</sub>O comprised the principal part of total calculated CO<sub>2</sub>-e emissions during spring and winter, while the contributions of CH<sub>4</sub> could not be ignored during summer and autumn. This study highlights the importance of seasonal N<sub>2</sub>O and CH<sub>4</sub> contributions, particularly during times of significant CH<sub>4</sub> consumption. For the accurate up-scaling of N<sub>2</sub>O and CH<sub>4</sub> fluxes to annual rates, a careful sampling design at site-level is required to capture the potentially considerable temporal and spatial variations of N<sub>2</sub>O and CH<sub>4</sub> emissions.

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## 1. Introduction

Nitrous oxide (N<sub>2</sub>O) and methane (CH<sub>4</sub>) are key radiatively active greenhouse trace gases which have been recognized to contribute global warming by 5% and 25%, respectively (Mosier, 1998). Quantification of the trace gases is a subject of great interest because accurate information is required to determine the contribution of these gases to global greenhouse gas fluxes (Khalil et al., 2002). IPCC (2007) has reported increased concentrations in N<sub>2</sub>O and CH<sub>4</sub> since industrial times, a concern since both gases, although present in lower concentrations to that of carbon dioxide (CO<sub>2</sub>), have 298 (N<sub>2</sub>O) and 25 (CH<sub>4</sub>) times the global warming potential of CO<sub>2</sub> over a 100-year time period. The current global



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atmospheric N<sub>2</sub>O and CH<sub>4</sub> concentrations are  $319 \pm 0.12$  ppb and 1774.62  $\pm$  1.22 ppb, respectively, and are increasing approximately 0.26% and 1.1% per year (IPCC, 2007).

Emission of N<sub>2</sub>O and CH<sub>4</sub> from various natural ecosystems has significant influences on the global climate change since they account for 44-54% of the total N<sub>2</sub>O emissions and 30-40% of the total CH<sub>4</sub> emissions, respectively (IPCC, 2007). Tropical soil and wetlands play an important role in the global carbon (C) and nitrogen (N) biogeochemical cycles, and are considered significant natural sources of N<sub>2</sub>O and CH<sub>4</sub>, contributing approximately 22-27% (N<sub>2</sub>O) and 24% (CH<sub>4</sub>), respectively, towards this inventory (Whalen, 2005). Coastal marsh ecosystem is characterized by high temporal and spatial variations involved with topographic feature, environmental factors and astronomic tidal fluctuation, and is very sensitive to global climate changes and human activities. Above all, the intertidal zone between terrestrial and aquatic coastal ecosystems may represent a high dynamic interface of intense material processing and transport, with potentially high decomposition and associated greenhouse gases emission (Hirota et al., 2007). In the past two decades, considerable efforts have been made to quantify the N<sub>2</sub>O and CH<sub>4</sub> fluxes in different coastal ecosystems, especially in estuarine salt marshes (Magalhães et al., 2007; Moseman-Valtierra et al., 2011), mangrove swamps (Allen et al., 2007; Ganguly et al., 2008), coastal lagoons (Gregorich et al., 2006; Hirota et al., 2007) and coastal marshes (Amouroux et al., 2002). In China, the studies on N<sub>2</sub>O and CH<sub>4</sub> emissions from coastal marshes started quite late (in the 2000s), and the related research primarily focused on the coastal tundra marshes in Antarctica (Sun et al., 2002; Zhu et al., 2008) and the salt marshes in the Yangtze River estuary (Yang et al., 2006; Wang et al., 2007) and Min River estuary (Zeng et al., 2010; Mou et al., 2012). However, information on the coastal marshes in other estuaries is still very scarce.

The Yellow River is well known as a sediment-laden river. Every year, approximately  $1.05 \times 10^7$  tons of sediment is carried to the estuary and deposited in the slow flowing landform, resulting in vast floodplain and special marsh landscape (Cui et al., 2009). Sediment deposition is an important process for the formation and development of coastal marshes in the Yellow River Delta. The deposition rate of sediment in the Yellow River not only affects the formation rate of coastal marsh, but also, to some extent, influences water or salinity status and plants succession. Coastal marsh is the main marsh type, with an area of 964.8 km<sup>2</sup>, accounting for 63.06% of total area of the Yellow River Delta (Cui et al., 2009). Suaeda salsa, an annual C<sub>3</sub> plant, is one of the most prevalent halophytes in the coastal marshes of the Yellow River estuary. As a pioneer plant, it has strong adaptations to environmental stresses, such as high salinity, flooding and sediment burial (Han et al., 2005). S. salsa generally germinates in late April, blooms in July, matures in late September and completely dies in late November. In the intertidal zone, three phenotypes are formed due to the differences of water and salinity in high marsh, middle marsh and low marsh. In recent years, the N and organic matter loadings of the Yellow River estuary have significantly increased due to the effects of human activities, and approximately 4650 tons of nutrient and  $4.33 \times 10^5$  tons of organic matter are discharged into Bohai Sea every year (Ocean Environmental Quality Communique of Shandong Province, 2009). Increases in N and organic matter loadings to estuarine and coastal environment can stimulate microbial processes and associated trace gases emission (Seitzinger and Kroeze, 1998). However, emissions of N<sub>2</sub>O and CH<sub>4</sub> from different coastal S. salsa marshes in the Yellow River estuary remains poorly documented till now. In addition, because microbial processes affecting trace gas production are regulated by many parameters including oxygen availability, temperature, water content, sediment redox potential, salinity, pH and microbially available C and N sources (Bauza et al., 2002; Whalen, 2005), evaluating the influences of different environmental factors on the emissions of  $N_2O$  and  $CH_4$  from coastal marsh will be of importance.

In this paper, we measured the  $N_2O$  and  $CH_4$  fluxes from the intertidal zone of the Yellow River estuary using the closed chamber technique. The aims of this study are: (i) to quantify and compare the  $N_2O$  and  $CH_4$  fluxes from different coastal *S. salsa* marshes and bare flat; (ii) to determine whether distinct spatial and temporal variation occurs in  $N_2O$  and  $CH_4$  flux throughout the day and in different seasons; (iii) to examine how environmental factors influence  $N_2O$  and  $CH_4$  emissions.

## 2. Materials and methods

### 2.1. Site description

The study was carried out in the intertidal zone of the Yellow River estuary, which is located in the Nature Reserve of Yellow River Delta (37°35′N-38°12′N, 118°33′E-119°20′E) in Dongying City, Shandong Province, China. The nature reserve is of typical continental monsoon climate with distinctive seasons. The temperature changes significantly during early spring and winter, and the freeze/thaw cycles frequently occur in topsoil in majority days, with the frozen depth ranged from 0 cm to 15 cm. The annual average temperature is 12.1 °C, and the frost-free period is 196 d. The average temperature in spring, summer, autumn and winter are 10.7 °C, 27.3 °C, 13.1 °C and -5.2 °C, respectively. The annual evaporation is 1962 mm; the annual precipitation is 551.6 mm, and about 70% of precipitation occurring between June and August. The soils are dominated by intrazonal tide soil and salt soil and the main vegetation types include Phragmites australis, S. salsa, Triarrhena sacchariflora, Tamarix chinensis and Imperata cylindrica.

Intertidal zone sediment is composed mainly of fine particles. Natural geomorphology and depositing zones are distinct. High marsh, middle marsh, and low marsh develop from the land to the sea. The high marsh is predominated by S. salsa (>90%) and P. australis (<10%), while middle marsh is predominated by S. salsa (>95%) and T. chinensis (<5%). Low marsh includes two distinct ecosystem-types. One is pure S. salsa community (100%), with sparse distribution in the intertidal zone, and the other is bare flat. The coverage and maximum aboveground biomass of S. salsa-P. australis, S. salsa-T. chinensis and S. salsa communities are 95%, 80%, 60% and 902.08  $\pm$  195.81 g m^{-2}, 564.89  $\pm$  99.66 g m^{-2}, 252.97  $\pm$ 29.24 g m<sup>-2</sup>, respectively. In this study, four typical sampling positions were laid in high S. salsa marsh (HSM), middle S. salsa marsh (MSM), low S. salsa marsh (LSM) and bare flat (BF) on the northern coastal marsh of the Yellow River estuary. The initial physical and chemical properties of topsoil (0-10 cm) in the four positions are shown in Table 1.

#### 2.2. Experimental design

Fluxes of N<sub>2</sub>O and CH<sub>4</sub> were measured by using static, manual stainless steel chambers and gas chromatography techniques. The chamber is an open-bottom a square box (50 cm  $\times$  50 cm  $\times$  50 cm) and equipped with an electric fan installed on the top wall of each chamber to make turbulence when chamber was closed. Outside of the chamber was covered with 2 cm thickness white foam to reduce the impact of direct radiative heating during sampling. In August 2009, the stainless steel base (50 cm  $\times$  50 cm  $\times$  20 cm) with a water groove on top was installed at the four sampling positions. During observations, the chamber was placed over the base filled with water in the groove to ensure airtightness, and the plant was covered within the chamber.

Sampling campaigns were undertaken during four seasons in October 2009 (autumn), December 2009 (winter), April 2010

Table 1

itiai pri	ysical and chemical	i properti	es" of top	DSO11 (U-	10 cm) in the four coastal m	harsnes in the Yellow R	aver estuary.			
Sites	Bulk density (g cm <sup>-3</sup> )	Grain composition (%)		tion (%)	Electrical conductivity	Water content	TC (%)	TN (%)	NH <sub>4</sub> <sup>+</sup> -N	$NO_3^ N$
		Clay	Silt	Sand	$(mS cm^{-1})$	$(\mathrm{cm}^{3} \mathrm{cm}^{-3})$			$(mg kg^{-1})$	$(mg kg^{-1})$
HSM	1.28 ± 0.03	8.00	51.46	40.54	15.45 ± 1.44	$0.353 \pm 0.007$	$1.22 \pm 0.02$	$0.039 \pm 0.001$	$2.26 \pm 0.71$	1.31 ± 0.39
MSM	$1.34 \pm 0.06$	8.67	49.67	41.66	13.84 ± 0.81	0.343 ± 0.005	$1.29 \pm 0.01$	$0.039 \pm 0.002$	3.62 ± 0.22	1.21 ± 0.27
LSM	1.29 ± 0.11	10.14	57.13	32.74	10.38 ± 2.08	0.347 ± 0.022	$1.27 \pm 0.02$	0.036 ± 0.001	$2.01 \pm 0.94$	$1.10 \pm 0.57$
BF	1.43 ± 0.01	13.10	73.09	13.81	10.85 ± 0.44	0.358 ± 0.017	$1.56 \pm 0.02$	$0.045 \pm 0.003$	$4.66 \pm 2.43$	$1.19 \pm 0.42$

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<sup>a</sup> Values are means (±S.E.); HSM, High Suaeda salsa marsh; MSM, Middle S. salsa marsh; LSM, Low S. salsa marsh; BF, Bare flat.

(spring) and July 2010 (summer). Each measurement campaign consisted of 12 chambers set up at four positions (three chambers per site): HSM and MSM were located near the shore and which were submerged only at high or middle tide, and LSM and BF were adjacent to the sea and which were frequently submerged at low tide. On each sampling date, measurements were conducted at 7:00, 9:30, 12:00, 14:30 and 17:00 h. Measurements representing different times of day were from sediments exposed during low tide. Four air samples inside the chamber were collected every 20 min over a 60 min period by using 100 ml syringe equipped with three-way stopcocks. Samples were injected into pre-evacuated packs, transported to the laboratory and analyzed within 36 h using gas chromatography (Agilent 7890A) equipped with FID and ECD. The gas chromatography configurations for analyzing N<sub>2</sub>O and CH<sub>4</sub> concentrations were according to the method of Wang and Wang (2003), and the fluxes calculation followed the description of Song et al. (2008).

#### 2.3. Environmental measurements

Air temperature and soil temperatures (0, 5, 10, 15 and 20 cm) were measured in each position during gas sampling. Soil volumetric moisture and electrical conductivity (EC) in 0-5 and 5-10 cm depths were determined in situ by high-precision moisture measuring instrument (AZS-2) and soil and solution EC meter (Field Scout), respectively. Soil moisture and EC were not determined in December 2009 since the topsoil (0–10 cm) was frozen. On each sampling date, two soil samples (0-10 cm) per site were taken for analyzing TC and TN contents by element analyzer (Elementar Vario Micro, German) and NH<sub>4</sub><sup>+</sup>-N and NO<sub>3</sub><sup>-</sup>-N contents by sequence flow analyzer (San<sup>++</sup> SKALAR, Netherlands).

#### 2.4. Conversion to CO<sub>2</sub>-equivalents

Global warming potentials of 298 for N<sub>2</sub>O and 25 for CH<sub>4</sub> over 100 years (IPCC, 2007) were used to convert N<sub>2</sub>O and CH<sub>4</sub> emissions to CO<sub>2</sub>-e for comparing their greenhouse impacts.

## 2.5. Statistical analysis

The results were presented as means of the replications, with standard error (S.E.). Statistical significance of differences at p < 0.05 between samples were analyzed using analysis of variance (ANOVA). Correlation analyses were used to examine the relationship between fluxes and the measured environmental variables. In all tests, differences were considered significantly only if p < 0.05.

## 3. Results

## 3.1. N<sub>2</sub>O fluxes from different coastal marshes

Over all sampling periods, N<sub>2</sub>O fluxes from the four positions ranged from  $-0.0147 \text{ mgN}_20 \text{ m}^{-2} \text{ h}^{-1}$  to  $0.0982 \text{ mgN}_20 \text{ m}^{-2} \text{ h}^{-1}$ , thus both emission and consumption of N<sub>2</sub>O occurred. Except for

MSM. N<sub>2</sub>O fluxes from the other positions (HSM, LSM and BF) differed significantly between seasons (p < 0.05) (Fig. 1). With the exception of HSM that were found to release N<sub>2</sub>O during all times of day and the seasons measured, the other positions showed consumptions in some sampling times and seasons. Significantly higher N<sub>2</sub>O emissions were observed in HSM compared to LSM during April 7:00 and 14:30 sampling, and for October 9:30, 17:00 and July 9:30 (p < 0.05). Moreover, N<sub>2</sub>O fluxes from HSM were significantly higher than those from MSM during April 14:30 and July 9:30 sampling (p < 0.05), or those from BF during July 9:30 sampling (p < 0.05). N<sub>2</sub>O fluxes also differed significantly among the four positions during spring (April), autumn (October) and winter (December) (p < 0.01), but not during summer (July) (p > 0.05) (Fig. 1). N<sub>2</sub>O fluxes from MSM were quite lower than those from other positions over all sampling periods, and the values from HSM were significantly higher than those from MSM and BF in spring, summer and autumn (p < 0.05) or those from LSM in spring and autumn (p < 0.01). Significantly higher N<sub>2</sub>O emission was observed in LSM in winter compared to MSM (p < 0.01) and BF (p < 0.05) (Fig. 1). The average N<sub>2</sub>O fluxes from the four positions during spring, summer, autumn and winter were 0.0325 mgN<sub>2</sub>  $O m^{-2} h^{-1}$ , 0.0089 mgN<sub>2</sub>O m<sup>-2</sup> h<sup>-1</sup>, 0.0119 mgN<sub>2</sub>O m<sup>-2</sup> h<sup>-1</sup> and  $0.0140 \text{ mgN}_20 \text{ m}^{-2} \text{ h}^{-1}$ , respectively, indicating that coastal marsh represented N<sub>2</sub>O emission across all the seasons sampled.

## 3.2. CH<sub>4</sub> fluxes from different coastal marshes

CH<sub>4</sub> fluxes from the four positions averaged between  $-0.7421 \text{ mgCH}_4 \text{ m}^{-2} \text{ h}^{-1}$  and  $0.4242 \text{ mgCH}_4 \text{ m}^{-2} \text{ h}^{-1}$  and had no significant differences between seasons (p > 0.05) (Fig. 2). During all times of day and the seasons measured, both emission and consumption of CH<sub>4</sub> occurred in the four positions, but the consumption was predominated. CH<sub>4</sub> fluxes from HSM were significantly different with those from LSM during April 7:00, 12:30, 14:30 and 17:00 sampling, and for July 7:00, October 14:30 and December 10:30 (p < 0.05). Significant differences were also observed between MSM and LSM during April 7:00, 12:30, 14:30 and July 12:30 sampling (p < 0.05), or between MSM and BF during April 14:30, July 14:30, October 7:00, 12:00 and December 7:00 sampling (p < 0.05). Over all sampling seasons, CH<sub>4</sub> fluxes from the four positions had no significant differences (p > 0.05) (Fig. 2). Moreover, CH<sub>4</sub> fluxes from MSM and LSM were generally higher than those from HSM and BF in some sampling times and seasons. The mean CH<sub>4</sub> fluxes from the four positions during spring, summer, autumn and winter were  $-0.0109 \text{ mgCH}_4 \text{ m}^{-2} \text{ h}^{-1}$ ,  $-0.0174 \text{ mgCH}_4 \text{ m}^{-2}$  $h^{-1}$ , -0.0141 mgCH<sub>4</sub> m<sup>-2</sup>  $h^{-1}$  and -0.0089 mgCH<sub>4</sub> m<sup>-2</sup>  $h^{-1}$ , respectively, indicating that coastal marsh acted as CH<sub>4</sub> sink across all the seasons sampled.

## 3.3. Global warming potential (GWP)

Across all the seasons sampled, the coastal marsh was a net source of N<sub>2</sub>O and a net sink of CH<sub>4</sub>. As N<sub>2</sub>O and CH<sub>4</sub> emissions were expressed as CO<sub>2</sub>-e emissions, N<sub>2</sub>O emissions contributed



**Fig. 1.** Nitrous oxide flux (mgN<sub>2</sub>O m<sup>-2</sup> h<sup>-1</sup>) from high *Suaeda salsa* marsh (HSM), middle *S. salsa* marsh (MSM), low *S. salsa* marsh (LSM) and bare flat (BF) in spring (April 2010), summer (July 2010), autumn (October 2009) and winter (December 2009) in the Yellow River estuary. Values are means (±S.E., *n* = 3).

47.29% and 20.45% of net CO<sub>2</sub>-e emissions measured during spring and winter (Table 2), which were much higher than the contributions of net CO<sub>2</sub>-e emissions measured during summer (14.42%) and autumn (17.84%). In contrast, CH<sub>4</sub> emissions contributed 27.09% and 35.18% of net CO<sub>2</sub>-e emissions measured during summer and autumn (Table 2), which were much higher than the contributions of net CO<sub>2</sub>-e emissions measured during spring (23.14%) and winter (14.58%). The contributions of CH<sub>4</sub> to the total calculated CO<sub>2</sub>-e emissions were significantly lower than those of N<sub>2</sub>O during spring and winter (p < 0.05), but not during summer and autumn (p > 0.05) (Table 2).

## 3.4. Environmental variables of coastal marsh

Total C, total N and NH<sub>4</sub>–N in the sediment of BF were generally higher than those in the other positions over all sampling periods (Table 3). Significant differences in total C between BF and other position (HSM, MSM or LSM), and total N between BF and HSM (or LSM) were observed (p < 0.05). No significant differences in NH<sub>4</sub>–N and NO<sub>3</sub>–N content were observed in the four positions (p > 0.05). During all times of day and the seasons measured, air temperatures and sediment temperatures did not differ between positions (p > 0.05). Although water contents in subsurface sediment (5–10 cm) were higher than those in surface sediment (0–5 cm), no significant differences were found among the four positions (p > 0.05). Both EC in surface and subsurface sediments (0–5, 5–10 cm) had significant differences among the four positions (p < 0.01), in the order of HSM > MSM > LSM > BF.

Pearson correlation analysis between  $N_2O$  (or  $CH_4$ ) fluxes and independent variables indicated that most of the correlations between  $N_2O$  (or  $CH_4$ ) fluxes and environmental parameters in the four positions were not significant (p > 0.05) (Table 4). In HSM, N<sub>2</sub>O fluxes showed significant positive correlations with 5, 10 and 20 cm sediment temperatures (p < 0.05), while CH<sub>4</sub> fluxes showed significant positive correlation with NH<sub>4</sub>–N (p < 0.05). CH<sub>4</sub> fluxes from MSM showed significant negative correlations with NO<sub>3</sub>–N and air temperature (p < 0.05), while those from LSM had significant negative correlation with 0 cm sediment temperature (p < 0.05). In BF, N<sub>2</sub>O fluxes had no significant correlations with environment parameters, while CH<sub>4</sub> fluxes showed significant positive correlations with 5, 10 and 20 cm sediment temperatures (p < 0.05). In addition, the correlations between environmental parameters determined during all times of day and N<sub>2</sub>O (or CH<sub>4</sub>) diurnal emissions were not significant (p > 0.05).

## 4. Discussion

## 4.1. Temporal variations of N<sub>2</sub>O and CH<sub>4</sub> fluxes

This study demonstrated that  $N_2O$  and  $CH_4$  emissions from coastal marshes varied significantly throughout different times of the day (Fig. 1, Fig. 2). Diurnal variations in trace gas flux have been reported in other coastal marsh studies. Lu et al. (1999) showed that  $CH_4$  fluxes from *Bruguiera sexangula* mangrove swamp sediment had large diurnal fluctuations, which were caused by the changes of tidal inundation rather than the changes of air or sediment temperatures. Zhu et al. (2008) indicated that  $N_2O$  emission from the coastal tundra marsh in eastern Antarctica greatly differed at different times of the day, with emission peaks coincided with the maximum of ground temperature. Significant diel cycle of  $N_2O$  emission was also observed in the mangrove (*Rhizophora* 



**Fig. 2.** Methane flux (mgCH<sub>4</sub> m<sup>-2</sup> h<sup>-1</sup>) from high *Suaeda salsa* marsh (HSM), middle *S. salsa* marsh (MSM), low *S. salsa* marsh (LSM) and bare flat (BF) in spring (April 2010), summer (July 2010), autumn (October 2009) and winter (December 2009) in the Yellow River estuary. Values are means (±S.E., n = 3).

Table 2
Estimated total N <sub>2</sub> O and CH <sub>4</sub> emission (mgCO <sub>2</sub> equivalents $m^{-2}$ ) <sup>a</sup> during the sampling periods in the coastal marshes of the Yellow River estuary

Items	Spring		Summer		Autumn		Winter	
	N <sub>2</sub> O	CH <sub>4</sub>	N <sub>2</sub> O	CH <sub>4</sub>	N <sub>2</sub> O	CH <sub>4</sub>	N <sub>2</sub> O	CH <sub>4</sub>
HSM	50297 ± 13189	-3568 ± 184	11390 ± 1978	-2700 ± 329	17770 ± 6340	2654 ± 2628	12425 ± 2651	102 ± 1105
MSM	3117 ± 1009	$-2410 \pm 1116$	3297 ± 1673	2716 ± 4445	4079 ± 2927	2904 ± 3211	3865 ± 2995	$-1246 \pm 1711$
LSM	4212 ± 2668	1316 ± 833	6959 ± 1178	648 ± 263	1744 ± 1920	1436 ± 2943	16529 ± 8552	$-2237 \pm 351$
BF	25273 ± 6868	1483 ± 1102	$3630 \pm 2409$	$-4384 \pm 1282$	$7684 \pm 4078$	$-11826 \pm 347$	3019 ± 2518	1378 ± 550
Coastal marsh	20725 ± 6634 (47.29%) <sup>b</sup>	$-795 \pm 774$ (23.14%) <sup>b</sup>	6319 ± 1264 (14.42%) <sup>b</sup>	$-930 \pm 1297$ (27.09%) <sup>b</sup>	7819 ± 2559 (17.84%) <sup>b</sup>	$-1208 \pm 2151$ (35.18%) <sup>b</sup>	8960 ± 2703 (20.45%) <sup>b</sup>	$-501 \pm 614$ (14.58%) <sup>b</sup>

HSM, High Suaeda salsa marsh; MSM, Middle S. salsa marsh; LSM, Low S. salsa marsh; BF, Bare flat.

<sup>a</sup> Estimates were derived by weighted averages of 60 date points from morning to late afternoon (7:00, 9:30, 12:00, 14:30, 17:00 h) for each sampling month. The IPCC (2007) conversion of N<sub>2</sub>O and CH<sub>4</sub> emissions to CO<sub>2</sub> equivalents is 298 and 25, respectively, over a 100-year timeframe.

<sup>b</sup> Percentage contribution of N<sub>2</sub>O or CH<sub>4</sub> during each season is given in parentheses.

*mangle*) swamp at Magueys Island, with emissions peaks coinciding with increased ammonium production (Bauza et al., 2002). Since environmental factors are involved directly with different microbe and plant activities, such as photosynthesis, aerobic and anaerobic respiration and nitrification–denitrification and are changeable in relative short term (diurnal) scale, the diurnal variations of N<sub>2</sub>O and CH<sub>4</sub> emissions from different coastal marshes are generally affected by different principal factors. However, our study showed that any environmental factors determined during all times of day had no significant correlation with N<sub>2</sub>O or CH<sub>4</sub> diurnal emissions. Similar result was drawn by Hirota et al. (2007) who found that any environmental factors seemed to have no effects on the diurnal variations of N<sub>2</sub>O and CH<sub>4</sub> flux under light condition. We considered that there were two probable reasons: (i)

the Yellow River estuary might be more complicated than those in other coastal marshes, and  $N_2O$  or  $CH_4$  diurnal emissions were controlled by multiple factors; (ii) since our study did not measure  $N_2O$  or  $CH_4$  fluxes and environmental factors over the complete tidal inundation cycles, the variations of some major factors during all times of night might be missed, which, to some extent, covered the main factors. Whether  $N_2O$  and  $CH_4$  fluxes undergo strong diel cycles and which factor controlled their diurnal cycle requires further study.

Seasonal variations in  $N_2O$  and  $CH_4$  emissions from coastal marshes were observed in our study (Fig. 1, Fig. 2) and also reported by others (Wang et al., 2007; Hirota et al., 2007; Song et al., 2008). Whalen (2005) observed that, in arctic, boreal and temperate regions, seasonal patterns of trace gas emission were governed by seasonal variability in temperatures affecting water

Table 3									
Environmental	parameters <sup>a</sup>	of coastal	marshes	over the	2009/	2010	sampling	camp	baign

Environmental parameters	HSM	MSM	LSM	BF
TC (%)	$1.29 \pm 0.06^{b}$	$1.49 \pm 0.18^{b}$	$1.35 \pm 0.08^{b}$	1.57 ± 0.11 <sup>c</sup>
TN (%)	$0.04 \pm 0.01^{b}$	$0.04 \pm 0.01^{\rm bc}$	$0.04 \pm 0.01^{b}$	$0.05 \pm 0.01^{\circ}$
$NH_4-N (mg kg^{-1})$	$2.39 \pm 1.06^{b}$	$2.90 \pm 0.68^{b}$	$2.57 \pm 0.67^{b}$	$3.09 \pm 0.88^{b}$
$NO_3-N$ (mg kg <sup>-1</sup> )	$1.49 \pm 0.50^{b}$	$1.91 \pm 0.93^{b}$	$1.31 \pm 0.97^{b}$	$1.18 \pm 0.40^{b}$
Air temperature (°C)	$15.32 \pm 14.82^{b}$	$14.57 \pm 14.07^{b}$	$13.39 \pm 12.94^{b}$	$14.28 \pm 13.37^{b}$
Sediment temperature (°C)				
0 cm	$18.44 \pm 16.68^{b}$	16.80 ± 14.73 <sup>b</sup>	12.56 ± 11.73 <sup>b</sup>	$15.00 \pm 14.76^{b}$
5 cm	18.33 ± 15.42 <sup>b</sup>	17.09 ± 14.17 <sup>b</sup>	13.47 ± 13.44 <sup>b</sup>	15.25 ± 14.59 <sup>b</sup>
10 cm	17.17 ± 14.57 <sup>b</sup>	16.44 ± 13.74 <sup>b</sup>	14.33 ± 12.68 <sup>b</sup>	14.23 ± 12.89 <sup>b</sup>
15 cm	16.15 ± 14.31 <sup>b</sup>	15.45 ± 12.72 <sup>b</sup>	14.14 ± 12.73 <sup>b</sup>	13.15 ± 12.11 <sup>b</sup>
20 cm	15.23 ± 12.11 <sup>b</sup>	14.68 ± 12.20 <sup>b</sup>	13.38 ± 12.38 <sup>b</sup>	13.07 ± 11.03 <sup>b</sup>
Water content (cm <sup>3</sup> cm <sup>-3</sup> )				
0–5 cm	$0.29 \pm 0.02^{b}$	$0.29 \pm 0.02^{b}$	$0.28 \pm 0.04^{b}$	$0.28 \pm 0.03^{b}$
5–10 cm	$0.39 \pm 0.01^{b}$	$0.38 \pm 0.03^{b}$	$0.39 \pm 0.01^{b}$	$0.39 \pm 0.01^{b}$
EC (mS cm <sup><math>-1</math></sup> )				
0–5 cm	18.91 ± 1.7 <sup>b</sup>	15.67 ± 3.59 <sup>c</sup>	$9.14 \pm 0.98^{d}$	$10.59 \pm 0.97^{\rm e}$
5–10 cm	$15.76 \pm 2.0^{b}$	11.37 ± 2.34 <sup>c</sup>	$8.04 \pm 1.09^{d}$	9.61 ± 1.20 <sup>e</sup>

HSM, High Suaeda salsa marsh; MSM, Middle S. salsa marsh; LSM, Low S. salsa marsh; BF, Bare flat.

<sup>a</sup> Values are means ( $\pm$ S.E.) of samples (n = 24 for TC, TN, NH<sub>4</sub>–N and NO<sub>3</sub><sup>-</sup>–N; n = 18 for water content and EC; n = 60 for air temperature and soil temperature) collected from high *S. salsa* marsh (HSM), middle *S. salsa* marsh (MSM), low *S. salsa* marsh (LSM) and bare flat (BF) over all sampling periods. Statistically significant differences among sampling positions (p < 0.05) were calculated for multiple comparisons using Kruskal–Wallis ANOVA and are indicated by different letters within each row.

Table 4
Pearson correlation analysis between $N_2O$ or $CH_4$ fluxes and environmental parameters <sup>a</sup> .

Environmental parameters	HSM		MSM	MSM		LSM		BF	
	N <sub>2</sub> O	CH <sub>4</sub>	N <sub>2</sub> O	CH <sub>4</sub>	N <sub>2</sub> 0	CH <sub>4</sub>	N <sub>2</sub> O	CH <sub>4</sub>	
тс	-0.237	0.088	-0.038	-0.881	0.573	-0.614	0.441	0.412	
TN	0.484	-0.051	-0.367	-0.814	-0.021	0.057	0.635	0.139	
NH <sub>4</sub> -N	-0.616	0.983 <sup>b</sup>	-0.092	0.293	0.445	-0.460	-0.741	-0.504	
NO <sub>3</sub> -N	-0.294	0.282	-0.524	$-0.984^{b}$	-0.062	-0.104	-0.525	0.497	
Air temperature	0.410	-0.059	0.042	$-0.488^{b}$	-0.124	-0.334	-0.270	0.263	
Sediment temperature									
0 cm	0.433	-0.004	-0.058	-0.345	-0.004	$-0.495^{b}$	0.163	0.319	
5 cm	0.536 <sup>b</sup>	-0.032	0.221	-0.235	-0.032	-0.233	0.269	0.479 <sup>b</sup>	
10 cm	0.565 <sup>b</sup>	-0.228	0.291	-0.093	-0.228	-0.294	-0.387	0.894 <sup>b</sup>	
15 cm	0.443	-0.374	0.313	-0.113	-0.374	-0.026	-0.022	0.348	
20 cm	0.657 <sup>b</sup>	-0.417	0.310	0.086	-0.016	0.017	-0.040	0.503 <sup>b</sup>	
Water content									
0–5 cm	-0.930	0.514	0.796	0.664	0.971	-0.996	-0.140	0.579	
5–10 cm	-0.644	0.876	-0.731	0.310	0.996	-0.972	0.420	0.929	
EC									
0–5 cm	-0.943	0.725	0.751	0.914	0.071	0.079	0.981	0.581	
5–10 cm	-0.848	0.306	0.862	0.679	-0.755	0.844	0.186	-0.539	

HSM, High Suaeda salsa marsh; MSM, Middle S. salsa marsh; LSM, Low S. salsa marsh; BF, Bare flat.

<sup>a</sup> Pair sample size, n = 4 for TC, TN, NH<sub>4</sub>-N and NO<sub>3</sub><sup>-</sup>-N; n = 3 for water content and EC; n = 20 for air temperature and soil temperature.

<sup>b</sup> p < 0.05. 95% Confidence level was given during Pearson correlation analysis.

availability, production of substrate precursors and microbial activity. However, N<sub>2</sub>O and CH<sub>4</sub> emissions from the coastal marshes in the Yellow River estuary seemed not to be affected by seasonal variability in temperatures though the estuary located in temperate region (37°35'N-38°12'N). For instance, N<sub>2</sub>O emissions from HSM and BF were the lowest in summer and the highest in spring. N<sub>2</sub>O fluxes from HSM and BF during spring were also higher than those during winter, while emissions from MSM and LSM during winter were higher than those during spring (Fig. 1). Moreover, CH<sub>4</sub> fluxes from MSM and LSM were positive during summer while those from HSM and BF were negative in the meantime (Fig. 2). We considered that the seasonal variations in  $N_2O$ and CH<sub>4</sub> emissions observed in this paper were probably related to the complex interactions of temperatures and other biotic/abiotic factors, such as water and salinity status (Whalen, 2005), plants (Hirota et al., 2007) and sediment substrate (Allen et al., 2007). Similar result was drawn by Hirota et al. (2007) who found that the temporal variations of N<sub>2</sub>O and CH<sub>4</sub> fluxes from coastal lagoon were mainly controlled by temperatures and aboveground biomass.

We found that high N<sub>2</sub>O emissions occurred during spring and winter and low fluxes occurred during summer and autumn. The results were different with others which showed that N2O emissions were high in summer and autumn and low in spring and winter (Wang et al., 2007; Søvik and Kløve, 2007). Many studies have indicated that, in the mid-high latitude and high altitude regions, freeze/thaw cycle occurred in late autumn, winter or early spring was a very important process to increase N<sub>2</sub>O production and emission since it could affect soil physical and biological properties greatly (Teepe et al., 2001; Song et al., 2008). As mentioned previously, the coastal marshes of the Yellow River estuary locate in the mid-latitude region and the freeze/thaw cycles frequently occur in topsoil in majority days during spring and winter (frozen depth ranged from 0 cm to 15 cm). Thus, the high N<sub>2</sub>O emissions occurred during spring and winter might be related to the frequent freeze/thaw cycles. Similar results were drawn by Song et al. (2008) who found significant N<sub>2</sub>O emissions from freshwater marsh in northeast China during the freeze/thaw cycle as the temperature increased in late spring. Although higher temperatures were observed in summer and autumn compared to spring and

winter, the N<sub>2</sub>O emissions were very low. The low N<sub>2</sub>O emissions during summer and autumn might be related to the variations and interactions of environmental factors in coastal marsh which concealed the significant effects of temperature. In this study, we considered that N<sub>2</sub>O emissions during summer and autumn were controlled by multiple factors, and among them, S. salsa, had significant influences. As mentioned previously, S. salsa is the most prevalent halophyte in the coastal marshes of the Yellow River estuary and it has strong adaptation to the tidal inundation due to the transportation mechanism of oxygen  $(O_2)$  from aboveground parts to roots by aerenchyma (Han et al., 2005). First, part of O<sub>2</sub> transported to the roots can be released to the rhizosphere soil, which produces oxidizing microenvironment around S. salsa rhizosphere (Han et al., 2005; Kong et al., 2008). Second, soil microbes in the rhizosphere have great effects on the turnovers of available C and N in rhizosphere soil (Toal et al., 2000). Wang et al. (2010) studied the rhizosphere effects of S. salsa, T. chinensis and I. cylindrica in the Yellow River estuary, and found that the microbe amount and microbial activity in rhizosphere soil were much higher than those in non-rhizosphere soil and significantly higher values were observed in S. salsa rhizosphere soil compared to T. *chinensis* and *I. cylindrica* (p < 0.01), which was more favorable for the turnovers of available C and N (Herman et al., 2006; Cheng, 2009). Third, the S. salsa roots can also excrete some small molecular compounds (glucide, organic acid, amino acid and phenolic compounds), which provides plentiful available C for the rhizosphere microbes (Cheng, 2009; Wang et al., 2010). Thus, the presence of S. salsa probably accelerated the nitrification- denitrification rate as soil microbes in the rhizosphere were supplied with plentiful organic C and proper amount of O<sub>2</sub>, and N<sub>2</sub>O might be reduced to N<sub>2</sub> by denitrification regardless of whether N<sub>2</sub>O was produced by nitrification or denitrification. In addition, the proper amount of O<sub>2</sub> around the rhizosphere could also be partly diffused to the non-rhizosphere soil though the topsoil was under anoxic conditions, which, to some extent, reduced N<sub>2</sub>O emissions. Since studies on the N<sub>2</sub>O absorption and emission of different plants have been widely reported (Yang and Chen, 1995; Chen et al., 2003), the low  $N_2O$  emission might also be linked to the  $N_2O$ absorption of S. salsa, which required to be verified.

We also found that the coastal marsh in the Yellow River estuary acted as CH<sub>4</sub> sink across all the seasons sampled, and higher CH<sub>4</sub> sinks occurred during summer and autumn than during spring and winter. As mentioned previously, the seasonal variations in CH<sub>4</sub> emissions from coastal marshes were probably affected by the complex interactions of temperatures and other biotic/abiotic factors. We considered that high CH<sub>4</sub> sinks occurred during summer and autumn, to some extent, were dependent on the interactions of thermal conditions and other abiotic factors (such as soil moisture, salinity and soil substrate), which might be more favorable for the activities of methanotrophs than methanogens. Moreover, the CH<sub>4</sub> sink formed in the coastal marsh depended on the equilibrium of CH<sub>4</sub> production and oxidation which were related to the variations and interactions of abiotic and biotic factors (Ding et al., 2004; Hirota et al., 2007). Among them, soil moisture and salinity had great effects on the production/consumption of CH<sub>4</sub> (Megonigal et al., 2003). In this study, we observed CH<sub>4</sub> consumption during all the seasons sampled although the soil moistures in sediment were high (0–5 cm, 0.28–0.29 cm<sup>3</sup> cm<sup>-3</sup>; 5–10 cm, 0.38–  $0.39 \text{ cm}^3 \text{ cm}^{-3}$ ) (Table 3). The result was contrary to mostly previous studies which described that CH<sub>4</sub> flux increased with higher moisture (Moore and Dalva, 1993; Ye et al., 2000), but was similar with the conclusion drawn by Hirota et al. (2007) who reported a negative partial correlation between CH<sub>4</sub> flux and soil moisture in the sandy shore of Lake Nakaumi. We considered that the negative impact of soil moisture on CH<sub>4</sub> emission was related to the interaction of moisture and salinity in sediment. Magenheimer

et al. (1996) showed that the CH<sub>4</sub> fluxes from a macrotidal salt marsh (Bay of Fundy) were inversely correlated ( $r^2 = 0.23$ , p = 0.001) with salinity of the upper porewater at the sampling site. Similar results were drawn by Chidthaisong and Conrad (2000) and Zeng et al. (2008) who found that high salinity inhibited the activities of methanogens or did harm to methanogens, which reduced CH<sub>4</sub> emission. In this study, the salinity (represented by EC) of the coastal marsh in the Yellow River estuary were high (0-5 cm, 9.14-18.91 mS cm<sup>-1</sup>; 5–10 cm, 8.04–15.76 mS cm<sup>-1</sup>) (Table 3), which might induce  $CH_4$  consumption. The sulfate  $(SO_4^{2-})$  concentrations in sediment substrate resulting from high SO<sub>4</sub><sup>2-</sup> content in seawater (Ivanou, 1992) were also considered dominant factor controlling CH<sub>4</sub> emissions from coastal marsh (Kreuzwieser et al., 2003). Because sulfate (SO<sub>4</sub><sup>2-</sup>) reducing bacteria (SRB) could compete H<sub>2</sub>/ CO<sub>2</sub> and ethylic acid (CH<sub>3</sub>COOH) with methanogen and the former had more strong affinity to the reaction substrates, the coexistence of SRB and  $SO_4^{2-}$  would inhibit the production and emission of  $CH_4$ (van der Gon et al., 2001). In this study, the  $SO_4^{2-}$  concentration (0.62-1.50%) (Fan et al., 2010) in the coastal marshes of the Yellow River estuary were very high and the anoxic condition in sediment resulting from high moisture could enhance the dissimilatory reduction of  $SO_4^{2-}$ , which would inhibit the CH<sub>4</sub> emission of S. salsa marsh. Similar results were drawn by Magenheimer et al. (1996) who also found the dominant role of  $SO_4^{2-}$  in inhibiting methanogenesis in salt-marsh sediments.

This study also showed that, across all the seasons sampled, the coastal marsh was a net source of N<sub>2</sub>O and a net sink of CH<sub>4</sub>. When N<sub>2</sub>O and CH<sub>4</sub> emissions were converted to CO<sub>2</sub>-e emissions, N<sub>2</sub>O comprised the principal part of total calculated CO<sub>2</sub>-e emissions, with largest emissions occurring during spring and winter (Table 2). Although this contrasted the standpoint proposed for Carex Lasiocarpa freshwater marsh ecosystem, where both N<sub>2</sub>O and CH<sub>4</sub> contributed the main part of total calculated CO2-e emissions measured during summer and autumn (Song et al., 2008), approximated the notion reported by Allen et al. (2007) who found that N<sub>2</sub>O dominated winter CO<sub>2</sub>-e emissions at both fringe and grey mangrove (Avicennia marina) positions when CH<sub>4</sub> emissions were lowest. This finding highlights the importance of adequately capturing seasonal variation of both N<sub>2</sub>O and CH<sub>4</sub> in the future in the coastal marshes of the Yellow River estuary, particularly during times of CH<sub>4</sub> consumption, where the high global warming potential of N<sub>2</sub>O can dominate trace gas emissions.

### 4.2. Spatial variations of N<sub>2</sub>O and CH<sub>4</sub> fluxes

In this study, we observed that the physical and chemical parameters of sediment differed in their magnitude among the four positions during all times of day and the seasons measured. Especially, significant differences in total C between BF and other position (HSM, MSM or LSM), total N between BF and HSM (or LSM), and EC among the four positions were observed (p < 0.05) (Table 4). Such differences would be due to the site-specific conditions such as topography, aspect, slope, hydrology and species composition which determine the magnitudes and variations of N<sub>2</sub>O or CH<sub>4</sub> at spatial scale (Allen et al., 2007; Hirota et al., 2007).

This study showed that N<sub>2</sub>O fluxes differed significantly among the four positions (p < 0.01) (Fig. 1), and the values from HSM were generally high and those from MSM were very low. Previous studies have indicated that temperatures had great effects on N<sub>2</sub>O emissions at spatial scale (Alongi et al., 2005; Gregorich et al., 2006). During all times of day and the seasons measured, although air temperatures and sediment temperatures did not differ between positions (p > 0.05), significant relationships still could be found in HSM. This indicated that thermal condition was an important factor affecting N<sub>2</sub>O emission across the coastal marsh, but its function might be covered by the interactions of other biotic or abiotic factors such as moisture, salinity, sediment substrate and vegetation. In this paper, we found that the moisture in topsoil (0–10 cm) was very close and had no significant difference among the four positions (Table 3), which were mainly correlated with the low spatial heterogeneity of sediment moisture and the strong evaporation of the coastal marsh. Over all sampling seasons, the coefficient of variation (CV) of sediment moisture at 0-5 and 5-10 cm depths in HSM, MSM, LSM and BF were 1.38%, 4.31%, 15.7%, 10.54% and 1.49%, 6.36%, 1.60%, 0.53%, respectively, indicating that the spatial variability of sediment moisture (0-5 or 5-10 cm) among the four positions was not very high. As mentioned previously, the annual evaporation in the Yellow River estuary is 1962 mm and the annual precipitation is 551.6 mm (evaporation/ precipitation ratio, 3.52), implying that the strong evaporation might cause insignificant difference of the topsoil moistures as the spatial variability of sediment moisture across the coastal marsh was not high and the measurements were carried out during low tide over a number of days. Because soil evaporation was mainly affected by surface sediment moisture and the evaporation of surface sediment (0-5 cm) was more vigorous than that of subsurface sediment (Yang et al., 1997), high sediment moisture was observed in subsurface sediment (5-10 cm) in our study (Table 3). We also found that both positive and negative impacts of moisture on N<sub>2</sub>O emissions occurred in salt marshes (Table 4), which were different with mostly previous studies that N<sub>2</sub>O emissions had negative correlation with moisture (Regina et al., 1996; Wang et al., 2005). We considered that N<sub>2</sub>O emissions in different positions depended on the fluctuation of soil moisture resulting from astronomic tide. For one thing, proper soil moisture contributes to a favorable aerobic-anerobic status for N<sub>2</sub>O production by denitrification or nitrification or both. For another, N<sub>2</sub>O may be transported from surface seawater to the salt marsh by tidal fluctuation (Senga et al., 2001). Comparatively, the EC in topsoil (0-10 cm) had significant difference among the four positions (Table 3), which was mainly related to the spatial variability of sediment texture. Nadler (1991) indicated that soil texture significantly affected the soluble capacity of salinity even if soil moisture was approximate, and high EC was generally observed in coarse texture soil (Lin et al., 2005). As mentioned above, the sediment moisture across the coastal marsh had no significant difference while the grain composition had high spatial variability (the CVs of clay, silt and sand across the coastal marsh were 22.71%, 18.42% and 40.01%, respectively) (Table 1), which probably resulted in significant difference of EC among the four positions. Because the strong evaporation in coastal marsh could cause considerable salinity to be accumulated in surface sediment (Fan et al., 2010) and high moisture in subsurface sediment was favorable for the transference of salinity from subsurface sediment to deep layers (Lin et al., 2005), high EC was observed in surface sediment (Table 3). This paper also indicated that the correlations (positive and negative) between EC and N<sub>2</sub>O emission were not significant (Table 4), which were different with mostly previous studies that N<sub>2</sub>O emissions had negative correlation with EC (Dalal et al., 2003; Wang et al., 2009). One possible explanation for positive effect was that the salinity in MSM, LSM and BF might not completely inhibit the N turnover in salt marsh and the activities of nitrifiers and denitrifiers in sediment (Lv et al., 2008). Site-level control of N<sub>2</sub>O emission was also attributed to the effects of nutrient status. Although significant differences in total C and total N were observed among the four positions, the positive/negative correlations between total C or total N and N<sub>2</sub>O emission were not significant (Table 4). Positive/negative relationships between N<sub>2</sub>O emissions and total C or total N have been reported in various studies (Allen et al., 2007; Søvik and Kløve, 2007) due to C/N regulations and interactions of other abiotic variables at spatial scale (Huang et al., 2002). NH<sub>4</sub>-N and NO<sub>3</sub>-N content mostly had negative correlations with N<sub>2</sub>O emission in the four

positions (Table 4), which was similar with the conclusions drawn by Huang et al. (2002) and Tauchnitz et al. (2008), but was contrary to mostly previous studies (Aelion et al., 1997; Muñoz-Hincapié et al., 2002). In HSM, MSM and LSM, one possible reason was related to the interactions of vegetation (S. salsa) and microorganism (nitrifiers and denitrifiers) that produced negative influences on the transformation of NH<sub>4</sub>-N or NO<sub>3</sub>-N during N<sub>2</sub>O production. Li et al. (2002) indicated that vegetation had great impacts on N<sub>2</sub>O emission through influencing the activities of soil microorganism. Because available C, NH<sub>4</sub>-N and NO<sub>3</sub>-N were important substrates participating in the processes of nitrification and denitrification (Tauchnitz et al., 2008), the production of N<sub>2</sub>O might be inhibited by the available substrates as they were significantly competed by both vegetation absorption and microorganism utilization (Li et al., 2002). In BF, one possible reason was correlated with the strong horizontal and vertical movement of NH<sub>4</sub>-N or NO<sub>3</sub>-N in surface sediment due to the effects of tide (Mou, 2010). Strong movement of NH<sub>4</sub>-N or NO<sub>3</sub>-N would decrease the available N participating in the processes of nitrification and denitrification, which was unfavorable for the production and emission of N<sub>2</sub>O. In addition, since the species composition, biomass and coverage of vegetation in the four positions were different as mentioned previously, the vegetation distributed continuously across the coastal marsh would be one of the key factors for the N<sub>2</sub>O emissions at spatial scale.

Although CH<sub>4</sub> fluxes from the four positions had no significant differences (p > 0.05), CH<sub>4</sub> emission patterns were different during all times of day and the seasons measured (Fig. 2). Significant negative relationships between CH<sub>4</sub> emission and air temperature in MSM (p < 0.05) and between CH<sub>4</sub> emission and 0 cm sediment temperature in LSM (p < 0.05) were observed, while significant positive correlations occurred between CH<sub>4</sub> emission and sediment temperatures (5, 10 and 20 cm) in BF (p < 0.05) (Table 4). These indicated that thermal condition was also an important factor affecting CH<sub>4</sub> emission across the coastal marsh, but its function might be covered in some coastal marshes by other biotic or abiotic factors. For spatial variations of CH<sub>4</sub> fluxes in coastal marsh, we considered that the differences of coverage, aboveground biomass and species composition were the main driving forces. Significantly lower CH<sub>4</sub> emissions occurred in BF compared to HSM, MSM and LSM due to the inexistence of vegetation (Fig. 2). The importance of vegetation for C fluxes in wetlands has been described in many studies (Joabsson et al., 1999; Hirota et al., 2006). Especially, it has been well known that wetland plants have complex gas transport system via their body and emit CH<sub>4</sub> from soil to the atmosphere (Schimel, 1995). Since the productions of CH<sub>4</sub> in sediment were very low as mentioned previously, CH<sub>4</sub> emissions from soil to atmosphere transported by vegetation were also very low. Moreover, the limited CH<sub>4</sub> production could also be partly or completely oxidized before it was transported to atmosphere. These indicated that the species composition, biomass and coverage of vegetation across the coastal marsh, to some extent, affected CH<sub>4</sub> emission patterns at spatial scale. Not only temporal but also spatial variations of CH<sub>4</sub> fluxes from coastal marsh were determined by soil moisture and salinity. As mentioned previously, although the formation of CH<sub>4</sub> sink in coastal marsh at temporal scale was negative influenced by soil moisture and salinity, both positive and negative effects of them on CH<sub>4</sub> fluxes were observed in different positions (Table 4), which induced the formation of different CH<sub>4</sub> emission patterns at spatial scale. Similar result was drawn by Hirota et al. (2007) who found that, in coastal ecosystems subjected to the fluctuation of soil moisture by astronomic tide, there would be both positive and negative impacts on CH<sub>4</sub> emissions. Site-level control of CH<sub>4</sub> emission was also related to the effects of nutrient status (Ding and Cai, 2002). In this study, both positive and negative correlations between CH<sub>4</sub> emission and nutrient status were observed

(Table 4). Especially, significantly higher relationships between  $CH_4$  emission and  $NH_4$ –N in HSM (p < 0.05) and between  $CH_4$  emission and  $NO_3$ –N in MSM (p < 0.05) occurred (Table 4). Hence, local nutrient differences due to topography, aspect, slope, hydrology and vegetation influenced the spatial differences of  $CH_4$  emissions at spatial level.

In order to better understand the differences of trace gas fluxes from the coastal marsh in the Yellow River estuary and up-scale trace gas emissions to annual inventories, our study highlights the need to strengthen the frequency of sampling *in situ* at longterm and different spatial scales, which identifies the magnitude of  $N_2O$  and  $CH_4$  variations at seasonal, diurnal and spatial scales, before values are up-scaled to annual inventories; and to further identify key drivers of trace gas emissions in coastal marsh to reveal the causalities of spatial and temporal variations.

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